

structure formation are mainly dictated by the nature and intensity of interactions between food biopolymers in the bulk and on the interfaces of the food system. The main low molecular weight components of food can affect food structure formation by interacting with biopolymers and acting on the charge, conformation, molecular weight, size and other physical-chemical properties of the food biopolymers as well as on thermodynamic quality of the aqueous medium thereby radically altering biopolymer interactions.

In this connection the objective of research is the elucidation of the effect of lipophilic molecules (models of the lipid and flavour food components) on the thermodynamic properties of proteins in binary and more complex aqueous solutions. Two lipophilic molecules of the same chain length, non polar decane and polar sodium decanoate were employed. The proteins investigated were ovalbumin and 11S globulin.

The thermodynamic parameters of the different types pair interactions (the second virial coefficients) were estimated using static light scattering data in the binary and ternary aqueous solutions of the biopolymers without and with lipophilic molecules. The limit of thermodynamic stability of the systems (spinodal curve) and the co-ordinates of the critical point were calculated. Experimental data were in good agreement with the calculated results.

The influence of lipophilic molecules on the conformational state of proteins in aqueous medium was characterized by differential scanning calorimetry. Comparison of the effect of unpolar and polar lipophilic molecules on the thermodynamic properties of proteins was carried out.

**Acknowledgment** — The research described in this paper was made possible in part by Grant No. MLNOOO from the International Science Foundation.

## PROTEIN/PROTEIN AND PROTEIN/SURFACTANT INTERACTIONS IN EMULSION SYSTEMS

JIANSHE CHEN and ERIC DICKINSON

*Procter Department of Food Science, University of Leeds, Leeds LS2 9JT, UK*

Mixed biopolymer interactions have been investigated in the bulk aqueous phase and in oil-in-water emulsion systems. The interactions between oppositely charged  $\beta$ -lactoglobulin and gelatin in aqueous phase is clearly observed in electrophoretic mobility measurement, where the calculated zeta potential is changed from approximately  $-12$  mV in pH 7.0 buffer solution to about  $-3$  mV in the presence of 0.8 wt% gelatin. The addition of gelatin to  $\beta$ -lactoglobulin stabilized emulsions (0.4 wt%  $\beta$ -lactoglobulin, 20 wt% n-hexadecane, 20 mM bis-tris-propane, pH 7.0) induces bridging flocculation at a suitable dosage of gelatin. The most favourable flocculation is observed at a gelatin concentration of 0.3 wt%, a concentration corresponding to approximately half coverage and a very small emulsion droplet electrophoretic mobility. Competitive adsorption experiments in emulsions containing mixed proteins show that the proteins are more strongly packed at the interface and less easy to displace in the presence of Tween 20. This trend is also confirmed by surface shear viscosity experiments. The addition of Tween 20 to 1-day-old mixed  $\beta$ -lactoglobulin + gelatin film reduces the surface shear viscosity

to lower steady values over a period of 24 h, whereas the introduction of Tween 20 to a 1-day-old individual  $\beta$ -lactoglobulin or gelatin film brings the surface shear viscosity down to zero in just a few minutes. We suppose that the negatively charged  $\beta$ -lactoglobulin and positively charged gelatin are strongly interacting with each other and so form a more closely packed and more highly cross-linked complex at the interface. This kind of complexed adsorbed protein film most probably has fewer interaction sites available for the non-ionic surfactant, and it may provide a diffusional barrier slowing down the rate of penetration of surfactant towards the oil-water interface. The experimental results suggest that there may be a synergistic effect occurring between two different kinds of protein at the oil-water interface which may be relevant to food emulsion stability in systems containing mixed food biopolymers.

## THE EFFECT OF COUNTER IONS ON THE PARTITIONING OF BIOMOLECULES IN UCON-DEXTRAN AND UCON-WATER SYSTEMS

HANS-OLOF JOHANSSON<sup>a</sup>, GUNNAR KARLSTRÖM<sup>b</sup> and FOLKE TJERNELD<sup>a</sup>

<sup>a</sup>Department of Biochemistry <sup>b</sup>Theoretical Chemistry, University of Lund, POB 124, S-22100 Lund, Sweden

Ucon-dextran is an aqueous polymer two-phase system, formed at room temperature, which can be used for protein partitioning (Harris *et al.*, 1991). Ucon is a water-soluble random copolymer of equal amounts of weight of ethylene oxide and propylene oxide. The observed partition coefficient of a charged protein in a Ucon-dextran system is an average value as a result of contributions from the partitioning of the single protein and its counter ions. Perchlorate and triethyl ammonium are two hydrophobic ions which have been used to direct the partitioning of albumin and lysozyme in Ucon-dextran systems.

In water solutions containing the hydrophobic Ucon-polymer a two-phase system is formed after temperature increase (Johansson *et al.*, 1993). This polymer-water system has been used to study the effect of the relative hydrophobicity of the amino acid side chains and different counter ions. Tryptophan (Trp) was found to be strongly partitioned to the hydrophobic polymer rich phase. The partitioning of Trp in a Ucon-water system can be directed to the polymer rich phase at low pH (pH 2 or less) with  $\text{NaClO}_4$  present in the system. At this pH the counter ion of Trp is  $\text{ClO}_4^-$ . At high pH (pH 10 or higher)  $\text{NaClO}_4$  will drive the partitioning of Trp to the water rich phase.  $\text{Na}^+$  is the counter ion of Trp in this case.  $\text{ClO}_4^-$  and  $\text{Na}^+$  have different affinities to the polymer rich phase which explains why Trp partitions differently at low or high pH. The effect of hydrophobic ions on Trp in Ucon-water systems is qualitatively the same for protein partitioning in Ucon-dextran systems.

## References

- Harris, P.A., Karlström, G. & Tjerneld, F. (1991). *Bioseparation*, **2**, 237–246.
- Johansson, H.-O., Karlström, G. & Tjerneld, F. (1993). *Macromolecules*, **26**, 4478–4483.